Sleep Loss Activates Cellular Inflammatory Signaling


Background: Accumulating evidence suggests that sleep disturbance is associated with inflammation and related disorders including cardiovascular disease, arthritis, and diabetes mellitus. This study was undertaken to test the effects of sleep loss on activation of nuclear factor (NF)-κB, a transcription factor that serves a critical role in the inflammatory signaling cascade.

Methods: In 14 healthy adults (seven women; seven men), peripheral blood mononuclear cell NF-κB was repeatedly assessed, along with enumeration of lymphocyte subpopulations, in the morning after baseline sleep, partial sleep deprivation (awake from 11 PM to 3:00 AM), and recovery sleep.

Results: In the morning after a night of sleep loss, mononuclear cell NF-κB activation was significantly greater compared with morning levels following uninterrupted baseline or recovery sleep, in which the response was found in female but not in male subjects.

Conclusions: These results identify NF-κB activation as a molecular pathway by which sleep disturbance may influence leukocyte inflammatory gene expression and the risk of inflammation-related disease.

Key Words: Inflammation, insomnia, nuclear factor-κB, sleep, sleep deprivation

Methods and Materials

Fourteen subjects (seven men and seven women) who were medically healthy, as determined by medical history, physical examination, and laboratory testing, were recruited between October 2006 and June 2007. All subjects fulfilled criteria for Never Mentally Ill as determined by Structured Clinical Interview for Diagnostic and Statistical Manual—IV (SCID) (mean age = 51.8 years [SD = 12.8]; 50% women; mean education level = 15.8 years [SD = 4.6]; mean body mass index [BMI] = 26.9 [SD = 3.6]; 35.7% were nonwhite). Females and males did not differ in mean age, education, BMI, or ethnicity (all p > .1). The sample focused on middle-aged and older adults, given the increased risk of inflammatory disorders with aging. Past or current Axis I DSM-IV psychiatric disorder, use of psychotropic medication, regular use of nonsteroidal antiinflammatory medications, and tobacco smoking were exclusionary. Subjects regularly slept between 10 PM and 7 AM, although use of actigraphy would have provided a more accurate assessment of sleep duration. The study was approved by the University of California at Los Angeles Institutional Review Board.

Subjects participated in a PSD protocol with acquisition of blood samples at 8:00 AM, given that PSD induces an increase of cellular markers of inflammation at this time point (see Supplement 1) (10). Nuclear extracts were prepared from isolated PBMC (12), and binding of activated NF-κB p65 to its consensus DNA sequence was measured using TransAM NF-κB p65 ELISA (Active Motif, Carlsbad, California; Supplement 1). Leukocyte cell subpopulations were enumerated in whole blood (Supplement 1).

Data were analyzed using SAS statistical software (version 9.13; SAS, Cary, North Carolina). To determine the effects of PSD on PBMC NF-κB activation and leukocyte subpopulations, repeated measures mixed-model analyses of variance were performed across the three conditions (baseline, PSD, recovery) covarying for age and sex. Both age and sex were covaried given a priori evidence that differences in proinflammatory cytokines are associated with these two factors. In addition, secondary analyses evaluated whether there were sex differences in the response of NF-κB. Significance level was set at p < .01.

Results

Subjects displayed an increase of PBMC NF-κB activation after PSD [condition effect: F(2,82) = 7.0, p < .01; Figure 1A]. Pairwise comparisons showed that levels of NF-κB activation were markedly

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increased the morning after PSD compared with baseline- and recovery morning levels (−.054, 95% confidence interval −.099 to −.009 ng NF-κB protein/μg total nuclear protein, \( p < .01 \); .057, .019−.165, 95% confidence interval, \( p < .01 \), with a 30% increase relative to baseline levels. Levels of NF-κB activation at baseline and recovery were similar (−.003, −.103−.035, 95% confidence interval, \( p = .68 \)). Secondary analyses examined whether females and males differed in response of NF-κB after PSD. As shown in Figure 1B, there was a significant sex \( \times \) condition interaction \( \left[ F(2,77.1) = 5.7 p < .01 \right] \), in which females showed an increase of PBMC NF-κB activation after PSD \( (p < .001) \) whereas males did not \( (p = .47) \). Finally, consistent with prior findings (6), PSD did not induce significant changes in leukocyte subpopulations; all \( ps > .1 \) (Table 1). Moreover, levels of NF-κB were not correlated with any of the leukocyte subpopulations including NK cell number (all \( ps > .1 \) by Spearman rho).

**Discussion**

Here we present evidence that acute sleep loss induces a rapid increase in activation of the transcription factor NF-κB in PBMC, providing a potential molecular mechanism for the effects of sleep loss on proinflammatory gene expression and circulating levels of inflammatory mediators (7, 8, 10). This observation that sleep loss results in nuclear translocation of NF-κB verifies previous bioinformatic indications (10) and closes an important gap in understanding the cellular mechanisms by which sleep loss enhances inflammatory biology in humans. Further studies that involve repeated measures of NF-κB are needed to evaluate the temporal dynamics of activation of this transcription factor across the diurnal period and in response to sleep loss. Nevertheless, these results are consistent with a previous study in *Drosophila* showing that sleep deprivation activates NF-κB (13).

Additional analyses showed that the effect of sleep loss on activation of the transcription factor NF-κB primarily occurs in females but not in males, although these results require cautious interpretation given the small sample size and their exploratory nature. Nevertheless, if sex differences in the response of NF-κB to sleep deprivation are confirmed, these results have implica-

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**Figure 1.** NF-κB DNA binding in peripheral blood mononuclear cells after baseline, partial night sleep deprivation, and recovery sleep in total sample (A) and in separate groups of females and males (B). NF, nuclear factor.
The mechanisms by which sleep disruption activates NF-κB were not evaluated in this study. PSD induces marked increases in cytokine and chemokine transcripts of norepinephrine are reported to be sufficient to result in a significant dose-dependent increase of NF-κB-binding activity in vitro (16). Whereas enhanced ex vivo expression of NF-κB following stress is due in part to the redistribution of leukocytes, PSD did not induce any detectable change in leukocyte subset distributions. Alternatively, sleep loss can induce increases of cortisol as well as IL-6 and TNF-α. However, PSD does not alter the temporal profile of cortisol (17), and cortisol at physiological doses is known to have potent anti-inflammatory effects by inhibiting NF-κB/Rel transcription factors and other proinflammatory signaling pathways (18). In contrast, sleep loss can induce expression of TNF-α, which is a potent inducer of NF-κB (11).

NF-κB activation is thought to contribute to the pathophysiology of diseases such as diabetes mellitus, cardiovascular disease, and atherosclerosis (16). Given evidence that sleep disturbance is associated with each of these medical disorders (10), sleep dependent NF-κB activation may be a common mechanism in the cumulative burden that finally leads to morbidity and mortality. Indeed, difficulty falling asleep or maintaining sleep is significantly associated with risk for nonfatal myocardial infarction or cardiovascular death, even after adjustment for multiple risk factors of coronary heart disease (19). Moreover, sleep loss upregulates several proinflammatory cytokines and chemokines, such as TNF-α, IL-1, IL-6, and IL-8, all of which are encoded by target genes of the IKK-β (inhibitor of NF-κB [IKB] kinase-β)-dependent NF-κB-activation pathway and are associated with tumor development and progression in humans and mice (4). Hence, these findings may also have implications for understanding associations between insomnia, disturbances in sleep–wake activity, and cancer in humans. Alternatively, activation of inflammatory pathways and increases of circulating IL-6, for example, have been found to be associated with insomnia and with disturbances in sleep onset (20), which together raise the possibility that targeting NF-κB may have therapeutic potential for insomnia.

Loss of sleep during only part of the night is one of the most common complaints of persons who experience environmental or psychological stress, travel across time meridians, engage in shift work, or have a psychiatric disorder. The results presented herein identify a key molecular pathway by which such sleep loss may influence immune system gene expression and inflammatory biology. These data should motivate further investigations to define the effects of recurrent sleep loss, as well as insomnia, on inflammatory mechanisms that underlie risk of cardiovascular and progression of chronic inflammatory disorders such as rheumatoid arthritis in humans. Finally, given that inflammation is a biologic consequence of aging, testing of interventions that target sleep might identify new strategies to constrain inflammation in older adults.

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Supplementary material cited in this article is available online.


Table 1. Leukocyte Subpopulations After Baseline, Partial-Night Sleep Deprivation, and Recovery Sleep

<table>
<thead>
<tr>
<th>Leukocyte Subpopulation (% of Cells)</th>
<th>Baseline Mean (SD)</th>
<th>PSD Mean (SD)</th>
<th>Recovery Mean (SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD 3 (T Lymphocyte)</td>
<td>67.9 (6.2)</td>
<td>67.4 (9.2)</td>
<td>62.2 (6.3)</td>
</tr>
<tr>
<td>CD 4 (T Helper)</td>
<td>44.7 (8.7)</td>
<td>44.4 (8.5)</td>
<td>39.4 (8.8)</td>
</tr>
<tr>
<td>CD 8 (Cytotoxic T Lymphocyte)</td>
<td>23.3 (5.5)</td>
<td>22.0 (6.0)</td>
<td>23.6 (6.1)</td>
</tr>
<tr>
<td>CD 16,56 (Natural Killer)</td>
<td>15.8 (6.3)</td>
<td>15.3 (6.1)</td>
<td>17.0 (5.3)</td>
</tr>
<tr>
<td>CD 19 (B Lymphocyte)</td>
<td>12.3 (4.8)</td>
<td>11.5 (4.1)</td>
<td>13.8 (3.7)</td>
</tr>
<tr>
<td>CD 14 (Monocyte)</td>
<td>19.9 (6.2)</td>
<td>19.6 (5.8)</td>
<td>19.8 (6.0)</td>
</tr>
</tbody>
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