α2-Adrenergic blockade mimics the enhancing effect of chronic stress on breast cancer progression

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Summary  Experimental studies in preclinical mouse models of breast cancer have shown that chronic restraint stress can enhance disease progression by increasing catecholamine levels and subsequent signaling of β-adrenergic receptors. Catecholamines also signal α-adrenergic receptors, and greater α-adrenergic signaling has been shown to promote breast cancer in vitro and in vivo. However, antagonism of α-adrenergic receptors can result in elevated catecholamine levels, which may increase β-adrenergic signaling, because pre-synaptic α2-adrenergic receptors mediate an autoinhibition of sympathetic transmission. Given these findings, we examined the effect of α-adrenergic blockade on breast cancer progression under non-stress and stress conditions (chronic restraint) in an orthotopic mouse model with MDA-MB-231HM cells. Chronic restraint increased primary tumor growth and metastasis to distant tissues as expected, and non-selective α-adrenergic blockade by phentolamine significantly inhibited those effects. However, under non-stress conditions, phentolamine increased primary tumor size and distant metastasis. Sympatho-neural gene expression for catecholamine biosynthesis enzymes

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1. Introduction

Clinical and epidemiological studies have shown that stress-related psychosocial factors in breast cancer patients can associate with accelerated progression of disease (Antoni et al., 2006; Chida et al., 2008). Experimental studies of stress effects in preclinical mouse models of breast cancer have shown that chronic restraint can increase the rate of primary tumor growth (Thaker et al., 2006) or cause primary tumor to become more metastatic (Sloan et al., 2010). Moreover, the non-selective β-blocking propranolol, abrogated the effect of chronic restraint on progression, strongly suggesting that stressor-induced catecholamines act through β-adrenergic receptors to mediate the effect of chronic restraint on the tumor microenvironment (Thaker et al., 2006; Sloan et al., 2010).

Catecholamines can also directly promote breast cancer cell proliferation in vitro through α-adrenergic receptors. Several α-adrenergic receptor antagonists blocked the proliferative effect of epinephrine and norepinephrine on human MCF-7 breast cancer cells in vitro (Vázquez et al., 1999). Similarly, an α-adrenergic receptor antagonist blocked the proliferative effect of epinephrine on human MDA-MB-231, human IBH-4, and mouse MC4-L5 breast cancer cells (Pérez Piñero et al., 2012).

Increased α-adrenergic signaling has also been shown to increase breast cancer progression in vivo under non-stress conditions with models that involve subcutaneous injection into the flank (Bruzone et al., 2008, 2009; Pérez Piñero et al., 2012) and an orthotopic model that primary mammary tumor and distant metastasis (Szpunar et al., 2013). In the latter orthotopic study (Szpunar et al., 2013), an α2-adrenergic agonist was shown to increase primary tumor growth rate and metastasis using the murine 4T1 cell line, which was found to lack functional α-adrenergic receptors. Thus, that study suggested that catecholamines may also signal host α-adrenergic receptors to promote breast cancer progression through indirect mechanisms.

Multiple reports have shown that pre-synaptic α2-adrenergic receptors can mediate an autoinhibition of sympathetic transmission (Hein et al., 1999; Scheinberger et al., 2001; Trendelenburg et al., 2001) and that disruption of this negative feedback signal with α2-adrenergic antagonists or gene inactivation of α2-adrenergic receptors enhances norepinephrine release (Starke, 2001). Thus, it is conceivable that blocking inhibitory α2-autoreceptors might increase sympa-tho-neural discharge and enhance β-adrenergic signaling with its known down-stream pro-tumor consequences for breast cancer progression (Thaker et al., 2006; Sloan et al., 2010; Madden et al., 2011).

Increased β-adrenergic signaling has been shown to increase accumulation of tumor-associated macrophages in primary mammary tumor, which increased angiogenesis and the expression of metastasis-promoting genes, including Tgfβ, Mmp9, and Vcam1 (Sloan et al., 2010). Moreover, direct β-adrenergic signaling in some breast cancer cell lines can increase expression of cytokines that promote tumor angiogenesis and metastasis, including vascular endothelial growth factor (VEGF) and interleukin-6 (IL-6) (Madden et al., 2011).

We were aware of no experimental in vivo studies that examined the effect of α-adrenergic blockade on breast cancer growing in an orthotopic tumor microenvironment. Given the reported ability of α-adrenergic antagonists to block direct effects of stress-typical catecholamines on breast cancer cell proliferation in vitro and the reported role for α-adrenergic signaling in breast cancer progression in vivo, we hypothesized that α-adrenergic blockade can inhibit breast cancer progression under non-stress and stress conditions in an orthotopic mouse model of breast cancer. However, given the reported ability of α-adrenergic receptors to mediate an autoimmune inhibition of sympathetic transmission, we also tested the alternate hypothesis that blockade of such α-autoreceptors can promote breast cancer progression under non-stress and stress conditions.

2. Methods

2.1. Animals

Seven-week-old female nu/nu mice (Charles River Laboratories, Crl:NU-Foxn1\(^{nu}\)) were housed under BSL2 barrier conditions on an individually ventilated cage (IVC) rack in dual filter disposable cages (Innovive, Inc.), in groups of 3–5 per cage, with corn cob bedding, nesting tissue, and ad libitum access to food and water on a 12:12 light:dark cycle at 22 °C. All procedures were carried out under protocols approved by the Institutional Animal Use and Care Committee of the University of California, Los Angeles.

2.2. Orthotopic breast cancer model

The MDA-MB-231HM cell line, a more highly metastatic derivative of the MDA-MB-231 human breast adenocarcinoma cell line (Chang et al., 2008), was transduced with a lentiviral vector containing firefly luciferase (Kaminskas et al., 2013) and cultured in DMEM with GlutaMAX™ and Pyruvate (Life Technologies-Gibco, #10569-044), supplemented with 10% FBS (Atlanta Biologicals, #S11550H), at
37 °C, in a 5% CO2 atmosphere. The tumor cells (2 × 10³) were injected into the left fourth mammary fat pad. Mammary tumor size was measured using digital calipers, and tumor volume was calculated as (length × width²)/2. The frequency and quantity of metastases were tracked in live mice by repeated noninvasive optical imaging of tumor-specific luciferase activity using the IVIS Lumina II system running Living Image 4.0 software (Caliper Life Sciences). After anesthetization with 2% isoflurane and intravenous injection of 150 mg/kg luciferin, mice were photographed under bright-field illumination and images were overlaid with luminescence data gathered over the maximum exposure period without pixel saturation (0.5–60 s). Metastasis was measured by triplicate determination at each time point of total bioluminescence in a region of interest of constant size located distant from the primary tumor (thoracic cavity where metastases appeared). Tissue-specific metastasis was measured ex vivo by bioluminescence immediately after sacrifice on day 28 or 32 of primary tumor growth.

2.3. Chronic restraint

Mice were randomly assigned to home cage control conditions or 2 h per day restraint for 20 days commencing 5 days before tumor cell inoculation. Mice were restrained in a confined space that prevented them from moving freely but did not press on them (Thaker et al., 2006). This paradigm has been shown to induce stress as shown by neuroendocrine activation (Thaker et al., 2006; Manni et al., 2008) and anxiety-like behaviors (Hermann et al., 1994).

2.4. α-Adrenergic receptor blockade

Phentolamine hydrochloride (Sigma, #P7547) in PBS vehicle, or an equivalent volume of vehicle alone for placebo in control mice and restraint-only mice, was injected subcutaneously at 5 mg/kg daily, commencing 5 days before tumor cell inoculation and continuing to the end of the experiment on day 28. Two experiments with phentolamine were conducted and included the following groups: (1) control; (2) restraint + placebo; (3) no restraint + phentolamine; (4) restraint + phentolamine. For each experiment, n = 3–5 mice per group. An additional experiment was conducted with phentolamine and the non-selective β-blocker, spropranolol (#P8688) at 2 mg/kg, to determine whether the promoting effects of phentolamine were mediated through β-adrenergic receptors and included the following groups: (1) control; (2) phentolamine; (3) propranolol; (4) phentolamine + propranolol (n = 5 mice per group). To examine the effect of selective α-adrenergic receptor blockade under non-stress conditions, an experiment was conducted with the α₁-blocker, prazosin hydrochloride (#P7791) at 2 mg/kg, and the α₂-blocker, efaxoxan hydrochloride (#E3263) at 5 mg/kg, and included the following groups: (1) control; (2) prazosin; (3) efaxoxan (n = 4–6 mice per group).

2.5. In vitro proliferation assays

Potential direct effects of phentolamine on MDA-MB-231HM proliferation in the absence and presence of catecholamines were tested by treating cells in vitro with phentolamine (0.0001–1 μM) and stressor-induced concentrations (Koolhaas et al., 2011) of epinephrine (0.0001, 0.001, and 0.01 μM; Sigma, #E4375) or norepinephrine (0.001, 0.01, and 0.1 μM; Sigma, #A9512). Proliferation was assayed by one-step MTS colorimetric assay (Promega, #G3852) at 24, 48, and 72 h after treatment initiation with a Thermo Scientific Multiskan MCC/340 microplate reader. Treatment media were replaced daily (DMEM with GlutaMAX, Pyruvate, 10% FBS). In each of two or three independent experiments, triplicate wells were prepared for each condition at each time point with initial seeding of 5000 cells per well.

2.6. Sympatho-adrenomedullary and sympatho-neural gene expression

To quantify gene expression of the key biosynthetic enzymes for catecholamine production in the adrenal medulla and sympathetic ganglia, total RNA was extracted from adrenal glands and from bi-lateral sympathetic chains of the thoracic T1–T10 vertebrae (Qiagen RNaseasy Mini Kits #74104 and #74704, respectively), with contaminating DNA cleared by on-column DNase digestion (Qiagen RNase-Free DNase Set, #79254). Resulting RNA was assayed by quantitative one-step RT-PCR (Qiagen Quantitite Probe RT-PCR, #204443) using sequence-specific primer-probe sets for biosynthetic enzymes in the adrenal medulla, i.e., tyrosine hydroxylase (Th), dopamine β-hydroxylase (Dbh), and phenylethanolamine N-methyltransferase (Pnmt), and in the sympathetic chain ganglia, i.e., Th, Dbh (Invitrogen-Applied Biosystems TaqMan Gene Expression Assays; Th, #Mm00447557_m1; Dbh, #Mm00460472_m1; Pnmt, #Mm00476993_m1). Transcripts for the catecholamine-degrading enzymes, monoamine oxidase A, Maa, and, monoamine oxidase B, Maob, were also assayed (Maa, #Mm00558004_m1; Maob, #Mm00555412_m1). RT-PCR was completed for tissue samples using triplicate determinations for each gene in each sample according to the manufacturer’s specified protocol on a Bio-Rad iCycler Real-Time Detection System, with enzyme threshold cycle values normalized to values of β-actin mRNA amplified in parallel (Actb, #Mm00607939_s1).

2.7. Statistical analysis

Data are presented as mean ± SE, and all statistical analyses were carried out using SPSS software (v.18). All distributions were examined for outliers and non-normality, and log transformations were applied to normalize distributions where necessary. Mixed-effects linear models (McCulloch et al., 2008) were used to analyze datasets pooled across experiments, where pharmacologic modification of longitudinal trajectory of breast cancer progression in vivo, or proliferation in vitro, was assessed by examining the group × time interaction term (i.e., restraint × phentolamine × day, catecholamine × phentolamine × day, or phentolamine × propranolol × day) and the group main effect alongside preplanned contrast analyses using the t-distribution. Composite catecholaminergic gene expression in the adrenal gland and in the sympathetic ganglia was assessed by examining the restraint × treatment interaction
α₂-Blockade mimics chronic stress in breast cancer progression

3. Results

3.1. Effects of α-adrenergic blockade on breast cancer progression under non-stress and stress conditions

To determine whether α-adrenergic blockade can inhibit breast cancer progression and attenuate the effect of chronic restraint stress on breast cancer progression, we measured the progression of MDA-MB-231HM human breast cancer in the mammary gland for 28 days and used in vivo optical imaging to quantify metastasis of the luciferase-tagged cells from the primary tumor. Chronic restraint increased the growth rate of primary tumor by 2.5-fold vs. controls (P = 0.025; Fig. 1A). α-Adrenergic blockade with phentolamine inhibited the effect of chronic restraint, as mean primary tumor volume in the chronic restraint plus α-adrenergic blockade group was not significantly different from controls (P values > 0.073; Fig. 1A). However, treatment with phentolamine in the absence of stress significantly increased primary tumor size compared to controls (P values < 0.05; Fig. 1A). Analysis of tumor weight on day 28 of primary tumor growth showed a similar pattern of results among groups (P values < 0.05; Fig. 1B).

Metastasis development correlated with primary tumor growth (Spearman correlation coefficient, r = 0.76, P < 0.0001). Chronic restraint significantly increased metastasis of primary breast tumor cells to distant tissues by an average of 4.5-fold vs. controls (P values < 0.001; Fig. 2A and B). The effect of chronic restraint was inhibited by α-adrenergic blockade, as metastasis values in the chronic restraint plus α-adrenergic blockade group were not significantly different from controls (P values > 0.714; Fig. 2A and B). However, α-adrenergic blockade in the absence of stress significantly increased metastasis levels compared to levels in control mice (P values < 0.01; Fig. 2A). Ex vivo analysis of tumor bioluminescence in lungs confirmed that both chronic restraint and α-adrenergic blockade by itself significantly increased metastasis (P values < 0.05; Fig. 2C and D).

3.2. Direct effects of α-adrenergic blockade on breast cancer cell proliferation

To determine whether the effects of α-adrenergic blockade in vivo were the result of direct signaling to tumor cells, we investigated the effect of drug signaling on tumor cell proliferation in vitro in the absence and presence of catecholamines. The effect of phentolamine on cell proliferation was not significantly different from untreated cells (P = 0.739; Suppl. Fig. S1A). Also, the combination of either epinephrine or norepinephrine with an equivalent concentration of phentolamine had no significant effect on cell proliferation (P values > 0.854; Suppl. Fig. S1B and C). These in vitro results suggested that the effects of α-adrenergic blockade in vivo under non-stress and stress conditions were likely due to indirect mechanisms.

3.3. Activation of the sympatho-adrenomedullary and sympatho-neural systems

To investigate the possibility that the effects of α-blockade under non-stress and stress conditions resulted from differential activity levels in catecholaminergic systems, we measured gene expression of the key biosynthetic enzymes for peripheral catecholamine production in the sympathoadrenomedullary and sympatho-neural systems. Tyrosine hydroxylase (Th) is the rate-limiting enzyme that initiates catecholamine synthesis; dopamine β-hydroxylase (Dbh) synthesizes norepinephrine; phenylethanolamine N-methyltransferase (Pnmt) converts norepinephrine to epinephrine. Chronic exposure to a variety of stressors increases expression of mRNA for Th, Dbh, and Pnmt in the adrenal medulla and Th and Dbh in sympathetic ganglia as a compensatory response to extensive catecholamine release during repeated stressful episodes (Kvetansky et al., 2009). Chronic restraint significantly increased adrenal gland catecholaminergic gene expression (composite of Th, Dbh, and Pnmt) by 37% vs. controls (P = 0.005, Fig. 3A). Although each of the component genes were increased by chronic restraint, Pnmt elevation appeared to be the most
sensitive (Th, 32%; Dbh, 28%; Pnmt, 47%; Fig. 3B). The effect of chronic restraint on catecholaminergic gene expression was not modified by α-adrenergic blockade (30% vs. controls, Fig. 3A; Th, 32%; Dbh, 31%; Pnmt, 26%; Fig. 3B), suggesting that the inhibitory effect of α-blockade on stress-enhanced cancer progression was mediated independently of adrenal gland-catecholamine activation. α-adrenergic blockade under non-stress conditions tended to increase adrenal catecholaminergic gene expression, but mean level did not reach significance (P = 0.112). We also measured mRNA expression of the catecholamine-degrading enzymes, monoamine oxidase A and B (Maoa, Maob). There was no significant effect of restraint on Maoa or Maob expression in the adrenal gland vs. controls (i.e., no restraint, no phentolamine) (P values > 0.485; Suppl. Fig. S2A and B). α-adrenergic blockade tended to increase adrenal gland Maoa expression in both the non-stress and stress groups vs. controls, but these levels did not reach significance (P values > 0.061; Suppl. Fig. S2A). α-adrenergic blockade also had no significant effect on adrenal gland Maob expression in the non-stress and stress groups vs. controls (P values > 0.708; Suppl. Fig. S2B).

In sympathetic ganglia, a different pattern of catecholaminergic gene expression occurred (composite of Th and Dbh, Fig. 3C). Here, chronic restraint had no significant effect on catecholaminergic gene expression (P = 0.483), and α-adrenergic blockade under stress conditions did not significantly alter catecholaminergic gene expression (P = 0.752).

However, α-adrenergic blockade under non-stress conditions increased catecholaminergic gene expression in sympathetic ganglia by 47% relative to controls (P = 0.054, Fig. 3C), suggesting that non-selective α-blockade may increase the release of sympatho-neural catecholamines that then act on β-adrenergic response pathways relevant to breast cancer progression. Individual gene expression for Th was more sensitive to non-selective α-blockade than Dbh (Th, 65%, Dbh, 34%, Fig. 3D). For gene expression of catecholamine-degrading enzymes in sympathetic ganglia, neither restraint by itself nor α-adrenergic blockade in either stress condition significantly affected Maoa or Maob expression (P values > 0.264; Suppl. Fig. S2C and D).

### 3.4. Effect of β-blockade on α-blockade-enhanced breast cancer progression

To further investigate the possibility that α-blockade mimicked chronic restraint enhancement of breast cancer progression by increasing sympatho-neural discharge and subsequent signaling of β-adrenergic receptors, we examined the effect of β-blockade on phentolamine-enhanced breast cancer progression. As shown in Fig. 4, there was a significant phentolamine × propranolol interaction effect on primary tumor growth (P = 0.006) and metastasis (P = 0.021). Propranolol inhibited the effect of phentolamine on tumor volume, as mean primary tumor volume in the phentolamine

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**Figure 2** Effect of α-adrenergic blockade on breast cancer metastasis to distant tissues under non-stress and stress conditions. (A) Distant metastasis was measured over 28 days, ***P < 0.001 for group effect. (B) Representative images of distant metastatic sites. (C) Ex vivo analysis of lung metastases, (a) P < 0.05 vs. control, (b) P < 0.05 vs. restraint + phentolamine. (D) Representative images of lungs showing tumor-specific bioluminescent signal. Y-axis: ×10^6 photons/s in (A) and (C).
plus propranolol group was significantly lower than in the phentolamine group ($P=0.001$; Fig. 4A). Likewise, propranolol inhibited the effect of phentolamine on distant metastasis compared to the phentolamine group ($P=0.001$; Fig. 4B and C).

### 3.5. Effects of selective $\alpha_1$- and $\alpha_2$-adrenergic blockade on breast cancer progression under non-stress conditions

To determine whether the effects of $\alpha$-blockade on breast cancer progression under non-stress conditions were mediated by a specific $\alpha$-adrenergic receptor subtype, we examined the effects of selective $\alpha_1$- and $\alpha_2$-blockers. Under non-stress conditions, the $\alpha_1$-blocker, prazosin, had no significant effect on primary tumor volume ($P=0.358$; Fig. 5A) or distant metastasis ($P=0.580$; Fig. 5B) compared to controls. However, the $\alpha_2$-blocker, efaxorphan, significantly increased both primary tumor volume ($P<0.001$; Fig. 5A) and distant metastasis ($P<0.001$; Fig. 5B).

### 4. Discussion

The $\alpha$-adrenergic antagonist, phentolamine, inhibited the promoting effect of chronic restraint stress on both primary tumor growth and metastatic dissemination in a preclinical orthotopic model of human breast cancer. However, under non-stress conditions, phentolamine increased primary tumor size and distant metastasis at a rate that was commensurate with the effect of chronic restraint. We found no evidence that phentolamine’s effects in vivo stemmed from direct effects on MDA-MB-231HM breast cancer cell proliferation. However, phentolamine caused a higher level of catecholaminergic gene expression in the peripheral sympatho-neural system compared to other groups. Moreover, the non-selective $\beta$-blocker propranolol inhibited the enhancing effects of phentolamine on breast cancer progression. We further found that the enhancing effects of non-selective $\alpha$-adrenergic antagonism on breast cancer progression were replicated with $\alpha_2$-blockade but not $\alpha_1$-blockade. These results suggest that $\alpha$-adrenergic signaling may play a significant role in breast cancer progression in vivo, but they also underscore the challenge that may lie in...
attempts to therapeutically interrupt that signaling dynamic with \(\alpha\)-adrenergic antagonism.

The present finding that \(\alpha_2\)-adrenergic blockade can mimic the enhancing effect of chronic stress on breast cancer progression is consistent with previous research in breast cancer models that indicate an increase in sympathetic activity can promote breast cancer progression by signaling through \(\beta\)-adrenergic receptors in the tumor microenvironment, which in turn can increase accumulation of tumor-associated macrophages, tumor angiogenesis, and the expression of metastasis-promoting genes (Thaker et al., 2006; Sloan et al., 2010). Several reports have shown that \(\alpha_2\)-adrenergic receptors mediate an autoinhibition of sympathetic transmission (Hein et al., 1999; Scheibner et al., 2001; Trendelenburg et al., 2001) and that disruption of this negative feedback signal with \(\alpha_2\)-adrenergic antagonists or gene inactivation of \(\alpha_2\)-adrenergic receptors enhances norepinephrine release (Starke, 2001). Subsequently, phentolamine, with affinity for both \(\alpha_1\) and \(\alpha_2\)-adrenergic receptors, has been shown to increase norepinephrine release from peripheral tissues ex vivo (Schelb et al., 2001). Consistent with that reported effect on norepinephrine by phentolamine, we found that phentolamine increased catecholaminergic gene expression in the peripheral sympatho-neural system and that phentolamine’s effects on breast cancer progression were mediated through \(\beta\)-adrenergic receptors. Moreover, we confirmed phentolamine’s \(\alpha_2\)-adrenergic effect by showing that the selective \(\alpha_2\)-antagonist, efaroxan, also significantly increased breast cancer progression but the selective \(\alpha_1\)-antagonist, prazosin, did not.

It is unclear why phentolamine’s \(\alpha_2\)-adrenergic antagonism did not increase sympatho-neural gene expression and breast cancer progression in mice that were also undergoing chronic restraint. Given that phentolamine is a reversible competitive antagonist (Dinsmore and Wyllie, 2008), it is possible that the drug exerted less total antagonism of pre-synaptic \(\alpha_2\)-adrenergic receptors because of increased competitive agonism by elevated systemic levels of catecholamine from the adrenal medulla in chronically
restrained mice. However, further research is needed to define these mechanisms.

Several observational studies have shown favorable outcomes for breast cancer patients using pharmacotherapy to block sympathetic signaling through β-adrenergic receptors (Powe et al., 2010; Melhem-Bertrandt et al., 2011; Barron et al., 2011; Botteri et al., 2013). The initial findings from the current preclinical study suggest the possibility that α-blockers exerting any α2-antagonism may increase sympathetic signaling and promote breast cancer progression through the β-adrenergic pathway. However, the inhibitory effect of phenolamine on cancer progression under stressful conditions in this study suggest that there may exist some circumstances in which α-blockers have potential clinical utility in breast cancer. Ultimately, more investigation in additional models of breast cancer is needed to test the generalizability of the current results. Also, further investigation of indirect mechanisms in the tumor microenvironment that likely mediate these results will determine whether such downstream mechanisms are similar to the ones found in previous studies (e.g., tumor-associated macrophage accumulation) and will allow for elucidation of the intracellular signaling pathways by which catecholamines regulate these pro-tumor effects. Given their seemingly paradoxical potential for help or harm in the context of breast cancer and the fact that many women take these drugs for conditions that include hypertension (Spatz et al., 2013), lower-urinary-tract symptoms (Boyd and Hilas, 2014), and anxiety symptoms (Raskind et al., 2013), we think careful investigation of α-blockers in this disease is further warranted.

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Conflict of interest

The authors report no conflict of interest.

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Appendix A. Supplementary data

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